

УДК 579.61:616.002.2,57.03

IL-8 СВЯЗЫВАЕТ ПУТИ NF-κB и Wnt/β-КАТЕНИН ПРИ ПЕРСИСТЕНТНОМ ВОСПАЛИТЕЛЬНОМ ОТВЕТЕ, ВЫЗВАННОМ ХРОНИЧЕСКОЙ ИНФЕКЦИЕЙ *Helicobacter pylori*¹

© 2023 г. L. Lin^a, *, B. Xie^b, J. Shi^c, C. M. Zhou^d, J. Yi^b, J. Chen^b, J. X. He^e, H. L. Wei^b, **

^aDepartment of Hematology & Oncology, Gansu Provincial Maternity and Child-Care Hospital,
Lanzhou, Gansu, 730050 China

^bKey Laboratory of Preclinical Study for New Drugs of Gansu Province, School of Basic Medical Sciences, Lanzhou University,
Lanzhou, Gansu, 730000 China

^cDepartment of Blood Transfusion, The Second Hospital of Lanzhou University,
Lanzhou, Gansu, 730000 China

^dDepartment of Clinical Laboratory Center, The First Hospital of Lanzhou University,
Lanzhou, Gansu, 730000 China

^eBasic Medical College, Gansu University of Chinese Medicine, Lanzhou, Gansu, 730000 China

*e-mail: gsfy_2021@sina.com

**e-mail: weihulai@lzu.edu.cn

Поступила в редакцию 03.08.2022 г.

После доработки 17.01.2023 г.

Принята к публикации 03.02.2023 г.

Инфекция *Helicobacter pylori* (*H. pylori*) иногда вызывает стойкий воспалительный ответ в эпителиальных клетках слизистой оболочки желудка человека, что может приводить к возникновению рака. Однако основной механизм канцерогенеза пока не выяснен. Нами разработаны модели хронической инфекции *H. pylori* в клетках GES-1 и на мышах C57BL/6J. Для определения уровня интерлейкина-8 (IL-8) использовали иммуоверментный анализ. Экспрессию мРНК и белков NF-κB p65, IL-8, Wnt2 и β-катенина определяли методами ПЦР в режиме реального времени, иммуноблотинга, иммунофлуоресцентного окрашивания и иммуногистохимии. Инфекцию *H. pylori* у мышей оценивали с помощью экспресс-теста на уреазу, окрашивания гематоксилином–эозином и серебрения по Вартину–Старри. Исследование морфологических изменений в слизистой оболочке желудка проводили методом электронной микроскопии. Выявлено, что в клетках слизистой оболочки желудка, инфицированных *H. pylori*, наряду с активацией сигнального пути NF-κB и повышением концентрации IL-8, значимо повышалась экспрессия Wnt2. На основании этих результатов можно предполагать, что IL-8 позитивно регулирует экспрессию гена *Wnt2*. При исследовании хронической инфекции *H. pylori* на модели мышей C57BL/6J показано, что у экспериментальных животных повышена частота предраковых поражений в ткани слизистой оболочки желудка. При сравнении ультраструктурных изменений в клетках слизистой оболочки желудка и анализе взаимосвязи между сигнальным путем NF-κB и экспрессией Wnt2 обнаружено, что инфекция *H. pylori* активирует сигнальные пути NF-κB, а массивное высвобождение IL-8 положительно коррелирует с высокой экспрессией белка Wnt2. Как следствие, активация сигнального пути Wnt/β-catenin может быть вовлечена в злокачественную трансформацию клеток слизистой оболочки желудка. Таким образом, хроническая инфекция *H. pylori* может приводить к персистентному воспалительному ответу: активировать путь NF-κB, способствовать высвобождению IL-8 и тем самым активировать путь Wnt/β-catenin. IL-8, по-видимому, играет роль линкера в сцеплении этих двух сигнальных путей.

Ключевые слова: *Helicobacter pylori*, интерлейкин-8, Wnt2, NF-κB, сигнальные пути, воспаление, рак желудка

DOI: 10.31857/S0026898423040134, **EDN:** QLUKEJ

СПИСОК ЛИТЕРАТУРЫ

1. Dvorak H.F. (1986) Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *New Engl. J. Med.* **315**, 1650–1659.
2. Medzhitov R. (2007) Recognition of microorganisms and activation of the immune response. *Nature*. **449**, 819–826.
3. Dias A., Garcia C., Majewski M., Wallner G., McCallum R.W., Poplawski C., Sarosiek J. (2011) Gastric

¹ Статья представлена авторами на английском языке.

- juice prostaglandins and peptide growth factors as potential markers of chronic atrophic gastritis, intestinal metaplasia and gastric cancer: their potential clinical implications based on this pilot study. *Dig. Dis. Sci.* **56**, 3220–3225.
4. Hussain S.P., Harris C.C. (2007) Inflammation and cancer: an ancient link with novel potentials. *Int. J. Cancer.* **121**, 2373–2380.
 5. Murata M. (2018) Inflammation and cancer. *Environ. Health Prevent. Med.* **23**, 50.
 6. (2020) Erratum: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **70**, 313.
 7. Vogtman E., Goedert J.J. (2016) Epidemiologic studies of the human microbiome and cancer. *Br. J. Cancer.* **114**, 237–242.
 8. Parsonnet J. (1994) Gastric adenocarcinoma and *Helicobacter pylori* infection. *Western J. Med.* **161**, 60.
 9. Valenzuela M.A., Canales J., Corvalan A.H., Quest A.F. (2015) *Helicobacter pylori*-induced inflammation and epigenetic changes during gastric carcinogenesis. *W. J. Gastroenterol.* **21**, 12742–12756.
 10. Clarke S.J., Chua W., Moore M., Kao S., Phan V., Tan C., Charles K., McMillan D.C. (2011) Use of inflammatory markers to guide cancer treatment. *Clin. Pharmacol. Ther.* **90**, 475–478.
 11. Pflug K.M., Sitcheran R. (2020) Targeting NF- κ B-inducing kinase (NIK) in immunity, inflammation, and cancer. *Int. J. Mol. Sci.* **21**(22), 8470.
 12. Taniguchi K., Karin M. (2018) NF- κ B, inflammation, immunity and cancer: coming of age. *Nat. Rev. Immunol.* **18**, 309–324.
 13. Cartwright T., Perkins N.D., Wilson C.L. (2016) NFKB1: a suppressor of inflammation, ageing and cancer. *FEBS J.* **283**, 1812–1822.
 14. Valovka T., Hottiger M.O. (2011) p65 controls NF- κ B activity by regulating cellular localization of I κ B β . *Biochem. J.* **434**, 253–263.
 15. Maeda S., Omata M. (2008) Inflammation and cancer: role of nuclear factor-kappaB activation. *Cancer Sci.* **99**, 836–842.
 16. Greten F.R., Eckmann L., Greten T.F., Park J.M., Li Z.W., Egan L.J., Kagnoff M.F., Karin M. (2004) IKK β links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell.* **118**, 285–96.
 17. Keates S., Hitti Y.S., Upton M., Kelly C.P. (1997) *Helicobacter pylori* infection activates NF- κ B in gastric epithelial cells. *Gastroenterology.* **113**, 1099–1109.
 18. Gambhir S., Vyas D., Hollis M., Aekka A., Vyas A. (2015) Nuclear factor kappa B role in inflammation associated gastrointestinal malignancies. *World J. Gastroenterol.* **21**, 3174–3183.
 19. Palena C., Hamilton D.H., Fernando R.I. (2012) Influence of IL-8 on the epithelial-mesenchymal transition and the tumor microenvironment. *Future Oncol.* **8**, 713–722.
 20. Visciano C., Liotti F., Prevete N., Cali G., Franco R., Collina F., de Paulis A., Marone G., Santoro M., Melillo R.M. (2015) Mast cells induce epithelial-to-mesenchymal transition and stem cell features in human thyroid cancer cells through an IL-8-Akt-Slug pathway. *Oncogene.* **34**, 5175–5186.
 21. Clevers H., Nusse R. (2012) Wnt/ β -catenin signaling and disease. *Cell.* **149**, 1192–205.
 22. Chiurillo M.A. (2015) Role of the Wnt/ β -catenin pathway in gastric cancer: an in-depth literature review. *World J. Exp. Med.* **5**, 84–102.
 23. Katoh M. (2003) WNT2 and human gastrointestinal cancer (review). *Int. J. Mol. Med.* **12**, 811–816.
 24. Kikuchi A., Yamamoto H., Kishida S. (2007) Multiplicity of the interactions of Wnt proteins and their receptors. *Cell. Signal.* **19**, 659–671.
 25. MacDonald B.T., Tamai K., He X. (2009) Wnt/ β -catenin signaling: components, mechanisms, and diseases. *Develop. Cell.* **17**, 9–26.
 26. Katoh M. (2001) Frequent up-regulation of WNT2 in primary gastric cancer and colorectal cancer. *Int. J. Oncol.* **19**, 1003–1007.
 27. Cheng X.X., Wang Z.C., Chen X., Sun Y., Kong Q.Y., Liu J., Gao X., Guan H.W., Li H. (2005) Frequent loss of membranous E-cadherin in gastric cancers: a cross-talk with Wnt in determining the fate of β -catenin. *Clin. Exp. Metastasis.* **22**, 85–93.
 28. Lin L., Wei H., Yi J., Xie B., Chen J., Zhou C., Wang L., Yang Y. (2019) Chronic CagA-positive *Helicobacter pylori* infection with MNNG stimulation synergistically induces mesenchymal and cancer stem cell-like properties in gastric mucosal epithelial cells. *J. Cell. Biochem.* **120**, 17635–17649.
 29. Keszei A.P., Goldbohm R.A., Schouten L.J., Jakszyn P., van den Brandt P.A. (2013) Dietary N-nitroso compounds, endogenous nitrosation, and the risk of esophageal and gastric cancer subtypes in the Netherlands Cohort Study. *Am. j. Clin. Nutr.* **97**, 135–146.
 30. Jakszyn P., Bingham S., Pera G., Agudo A., Luben R., Welch A., Boeing H., Del Giudice G., Palli D., Saieva C., Krogh V., Sacerdote C., Tumino R., Panico S., Berglund G., Siman H., Hallmans G., Sanchez M.J., Larranaga N., Barricarte A., Chirlaque M.D., Quiros J.R., Key T.J., Allen N., Lund E., Carneiro F., Linseisen J., Nagel G., Overvad K., Tjonneland A., Olsen A., Bueno-de-Mesquita H.B., Ocke M.O., Peeters P.H., Numans M.E., Clavel-Chapelon F., Trichopoulou A., Fenger C., Stenling R., Ferrari P., Jenab M., Norat T., Riboli E., Gonzalez C.A. (2006) Endogenous versus exogenous exposure to N-nitroso compounds and gastric cancer risk in the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST) study. *Carcinogenesis.* **27**, 1497–1501.
 31. Ah-Koon L., Lesage D., Lemadre E., Souissi I., Fagard R., Varin-Blank N., Fabre E.E., Schischmanoff O. (2016) Cellular response to alkylating agent MNNG is impaired in STAT1-deficient cells. *J. Cell. Mol. Med.* **20**, 1956–1965.
 32. Zhang C., Cai T., Zeng X., Cai D., Chen Y., Huang X., Gan H., Zhuo J., Zhao Z., Pan H., Li S. (2018) Astragaloside IV reverses MNNG-induced precancerous lesions of gastric carcinoma in rats: regulation on glycolysis through miRNA-34a/LDHA pathway. *Phytother. Res.* **32**, 1364–1372.

33. Yan Z. ., Xu T.T., An Z.T., Hu Y., Chen W.Z., Zhu F.S. (2018) Injury of human gastric epithelial GES-1 cells by MNNG and its effects on Wnt/beta-catenin signaling pathway. *Sheng Li Xue Bao* (in Chinese). **70**, 262–268.
34. Tsukamoto T., Mizoshita T., Tatematsu M. (2007) Animal models of stomach carcinogenesis. *Toxicol. Pathol.* **35**, 636–648.
35. Alm R.A., Trust T.J. (1999) Analysis of the genetic diversity of *Helicobacter pylori*: the tale of two genomes. *J. Mol. Med.* **77**, 834–846.
36. Zhang Y., Sun H., Chen X., Li J., Zhao H., Geng L., Li B. (2016) Functional profile of gastric epithelial cells infected with *Helicobacter pylori* strains. *Microb. Pathog.* **95**, 77–81.
37. Wang L., Zhou Y., Peng J., Zhang Z., Jiang D.J., Li Y.J. (2008) Role of endogenous nitric oxide synthase inhibitor in gastric mucosal injury. *Can. J. Physiol. Pharmacol.* **86**, 97–104.
38. Guo X.B., Guo L., Zhi Q.M., Ji J., Jiang J.L., Zhang R.J., Zhang J.N., Zhang J., Chen X.H., Cai Q., Li J.F., Yan M., Gu Q.L., Liu B.Y., Zhu Z.G., Yu Y.Y. (2011) *Helicobacter pylori* induces promoter hypermethylation and downregulates gene expression of IRX1 transcription factor on human gastric mucosa. *J. Gastroenterol. Hepatol.* **26**, 1685–1690.
39. Nurmi A., Vartiainen N., Pihlaja R., Goldsteins G., Yrjanheikki J., Koistinaho J. (2004) Pyrrolidine dithiocarbamate inhibits translocation of nuclear factor κ-B in neurons and protects against brain ischaemia with a wide therapeutic time window. *J. Neurochem.* **91**, 755–765.
40. Zhang B., Xu J., Quan Z., Qian M., Liu W., Zheng W., Yin F., Du J., Zhi Y., Song N. (2018) Klotho protein protects human keratinocytes from UVB-induced damage possibly by reducing expression and nuclear translocation of NF-κB. *Med. Sci. Monit.* **24**, 8583–8591.
41. Fernandes J.C., Martel-Pelletier J., Pelletier J.P. (2002) The role of cytokines in osteoarthritis pathophysiology. *Biorheology.* **39**, 237–246.
42. Monick M.M., Aksamit T.R., Geist L.J., Hunninghake G.W. (1994) Dexamethasone inhibits IL-1 and TNF activity in human lung fibroblasts without affecting IL-1 or TNF receptors. *Am. J. Physiol.* **267**, L33–L38.
43. Seidel P., Merfort I., Hughes J.M., Oliver B.G., Tamm M., Roth M. (2009) Dimethylfumarate inhibits NF-κB function at multiple levels to limit airway smooth muscle cell cytokine secretion. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **297**, L326–L339.
44. Liu A.Q., Xie Z., Chen X.N., Feng J., Chen J.W., Qin F.J., Ge L.Y. (2017) Fas-associated factor 1 inhibits tumor growth by suppressing *Helicobacter pylori*-induced activation of NF-κB signaling in human gastric carcinoma. *Oncotarget.* **8**, 7999–8009.
45. Shalapour S., Karin M. (2015) Immunity, inflammation, and cancer: an eternal fight between good and evil. *J. Clin. Investig.* **125**, 3347–3355.
46. David J.M., Dominguez C., Hamilton D.H., Palena C. (2016) The IL-8/IL-8R axis: a double agent in tumor immune resistance. *Vaccines.* **4**(3), 22.
47. Perugorria M.J., Olaizola P., Labiano I., Esparza-Baquer A., Marziani M., Marin J.J.G., Bujanda L., Banales J.M. (2019) Wnt-beta-catenin signalling in liver development, health and disease. *Nat. Rev. Gastroenterol. Hepatol.* **16**, 121–136.
48. Basu S., Cheriyaundath S., Ben-Ze'ev A. (2018) Cell-cell adhesion: linking Wnt/β-catenin signaling with partial EMT and stemness traits in tumorigenesis. *F1000Res.* **7**, 1488.

IL-8 Links NF-κB and Wnt/β-Catenin Pathways in Persistent Inflammatory Response Induced by Chronic *Helicobacter pylori* Infection

L. Lin^{1, *}, B. Xie², J. Shi³, C. M. Zhou⁴, J. Yi², J. Chen², J. X. He⁵, and H. L. Wei^{2, **}

¹Department of Hematology & Oncology, Gansu Provincial Maternity and Child-Care Hospital, Lanzhou, Gansu, 730050 China

²Key Laboratory of Preclinical Study for New Drugs of Gansu Province, School of Basic Medical Sciences, Lanzhou University, Lanzhou, Gansu, 730000 China

³Department of Blood Transfusion, The Second Hospital of Lanzhou University, Lanzhou, Gansu, 730000 China

⁴Department of Clinical Laboratory Center, The First Hospital of Lanzhou University, Lanzhou, Gansu, 730000 China

⁵Basic Medical College, Gansu University of Chinese Medicine, Lanzhou, Gansu, 730000 China

*e-mail: gsfy_2021@sina.com

**e-mail: weihulai@lzu.edu.cn

Helicobacter pylori (*H. pylori*) infection can cause persistent inflammatory response in human gastric mucosal epithelial cells, which may result in the occurrence of cancer. However, the underlying mechanism of carcinogenesis has not been elucidated yet. Herein, we established the models of chronic *H. pylori* infection in GES-1 cells and C57BL/6J mice. Interleukin 8 (IL-8) level was detected by ELISA. The expression of NF-κB p65, IL-8, Wnt2 and β-catenin mRNA and proteins was evaluated by real-time PCR, Western blotting, immunofluorescence staining, and immunohistochemistry. The infection of *H. pylori* in mice was evaluated by rapid urease test, H&E staining and Warthin–Starry silver staining. The morphological changes of gastric mucosa were observed by electron microscopy. Our results showed that in *H. pylori* infected gastric mucosal cells along with activation of NF-κB signaling pathway and increase of IL-8 level, the expression of Wnt2 was

also increased significantly, which preliminarily indicates that IL-8 can positively regulate the expression of *Wnt2*. Studies in chronic *H. pylori* infected C57BL/6J mice models showed that there was an increased incidence of premalignant lesions in the gastric mucosa tissue. Through comparing changes of gastric mucosal cell ultrastructure and analyzing the relationship between NF- κ B signaling pathway and *Wnt2* expression, we found that *H. pylori* infection activated NF- κ B signal pathways, and the massive release of IL-8 was positively correlated with the high expression of *Wnt2* protein. Subsequently, the activated Wnt/ β -catenin signal pathways may be involved in the malignant transformation of gastric mucosal cells. Collectively, *H. pylori* chronic infection may continuously lead to persistent inflammatory response: activate NF- κ B pathway, promote IL-8 release and thereby activate Wnt/ β -catenin pathway. IL-8 probably plays an important role of a linker in coupling these two signal pathways.

Keywords: *Helicobacter pylori*, interleukin 8, *Wnt2*, NF- κ B, signaling pathway, inflammation, gastric cancer